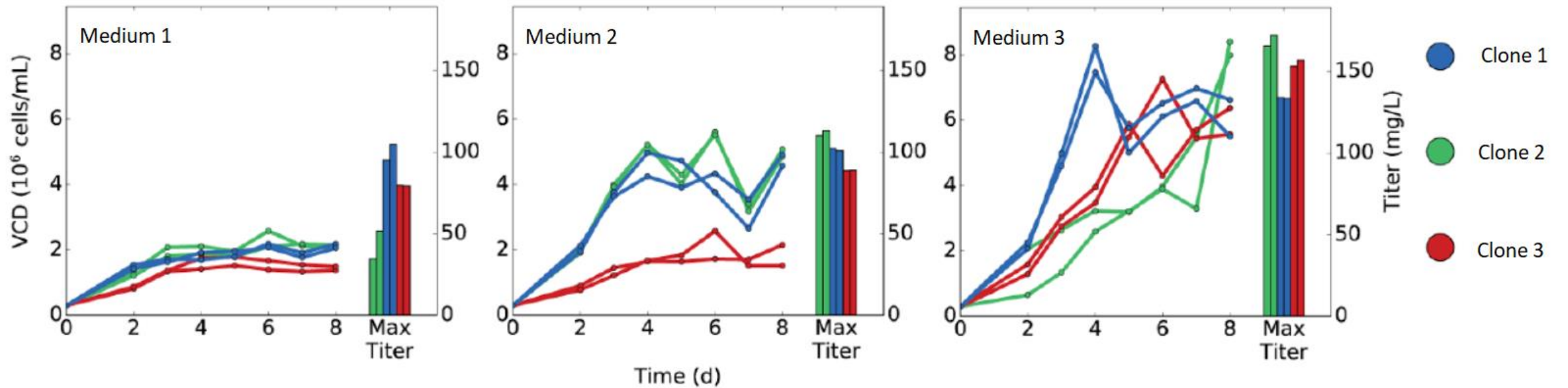


RESOURCES FOR METABOLIC MODELING IN MAMMALIAN CELL CULTURE

Nathan E. Lewis, PhD
Depts of Pediatrics and Bioengineering
University of California, San Diego
May 5, 2019

MODULATING CELL METABOLISM: THE EASIEST WAY TO IMPACT CELL GROWTH AND TITER



HOW INDIVIDUAL METABOLITES IMPACT MAMMALIAN CELL CULTURE

Necessary nutrients

Metabolites providing biomass precursors

- Sugars
- Amino acids

Metabolites impacting enzyme function

- Vitamins and cofactors
- Metal ions

Additives that do other things

- Hormones
- Anti-foam

Detrimental metabolites

Substrate competition

- Imbalanced amino acids competing for transporters
- Sugar nucleotides competing for glycan substrates

Toxic molecules inhibiting growth

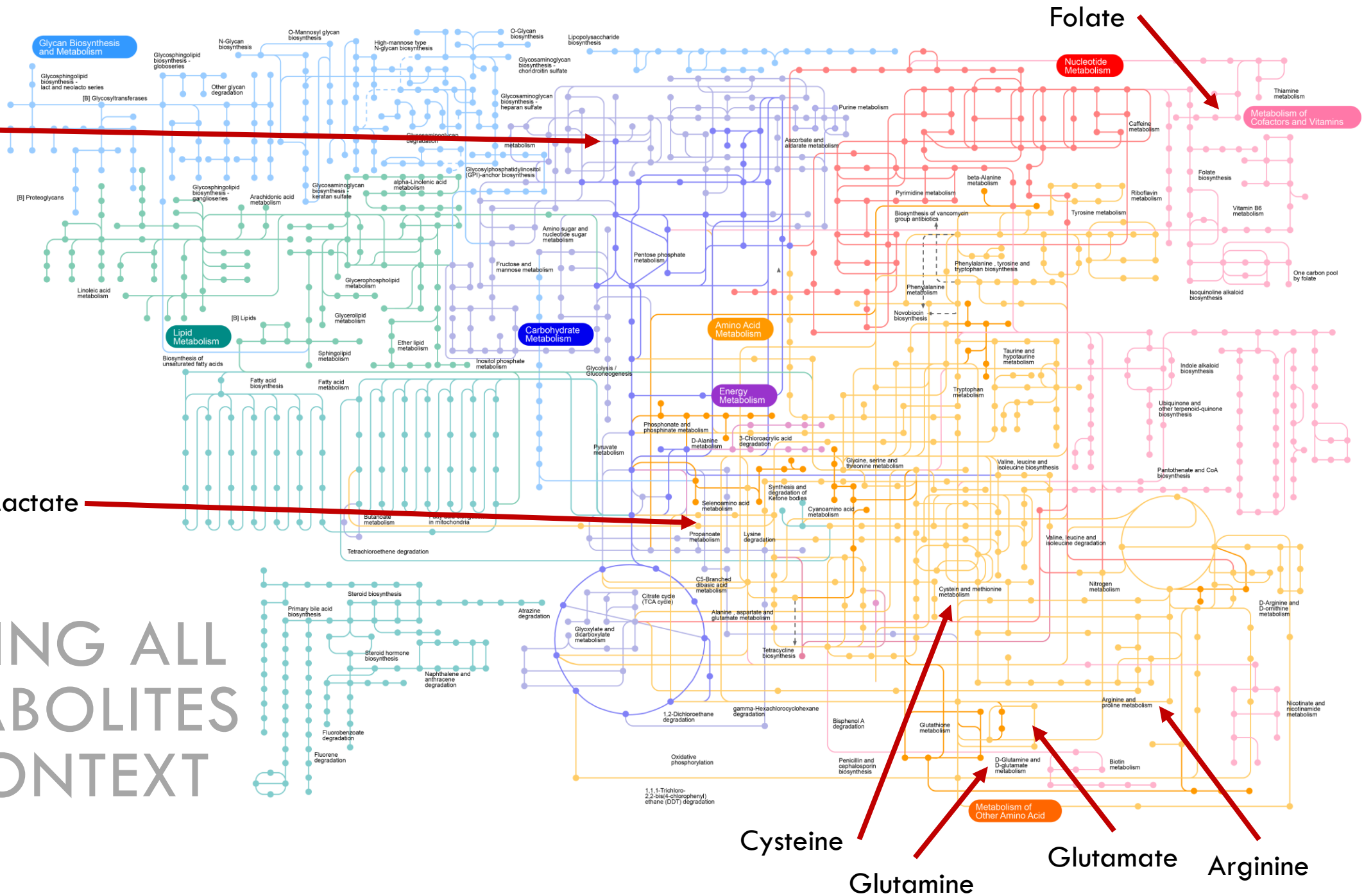
- Lactate
- Ammonia
- Excess salts and other osmolites
- Amino acid pathway intermediates

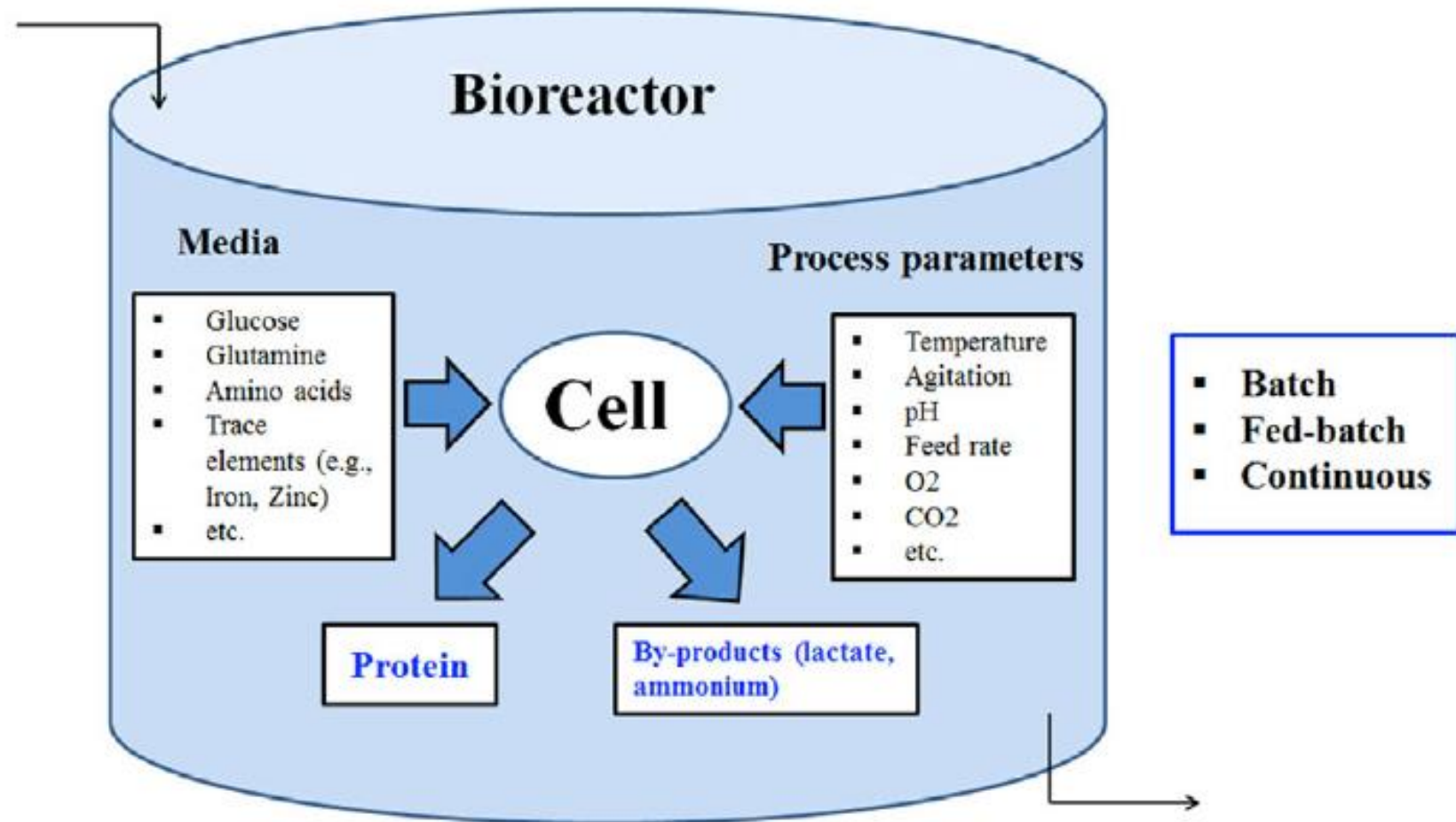
Glucose

Folate

Lactate

PUTTING ALL
METABOLITES
IN CONTEXT

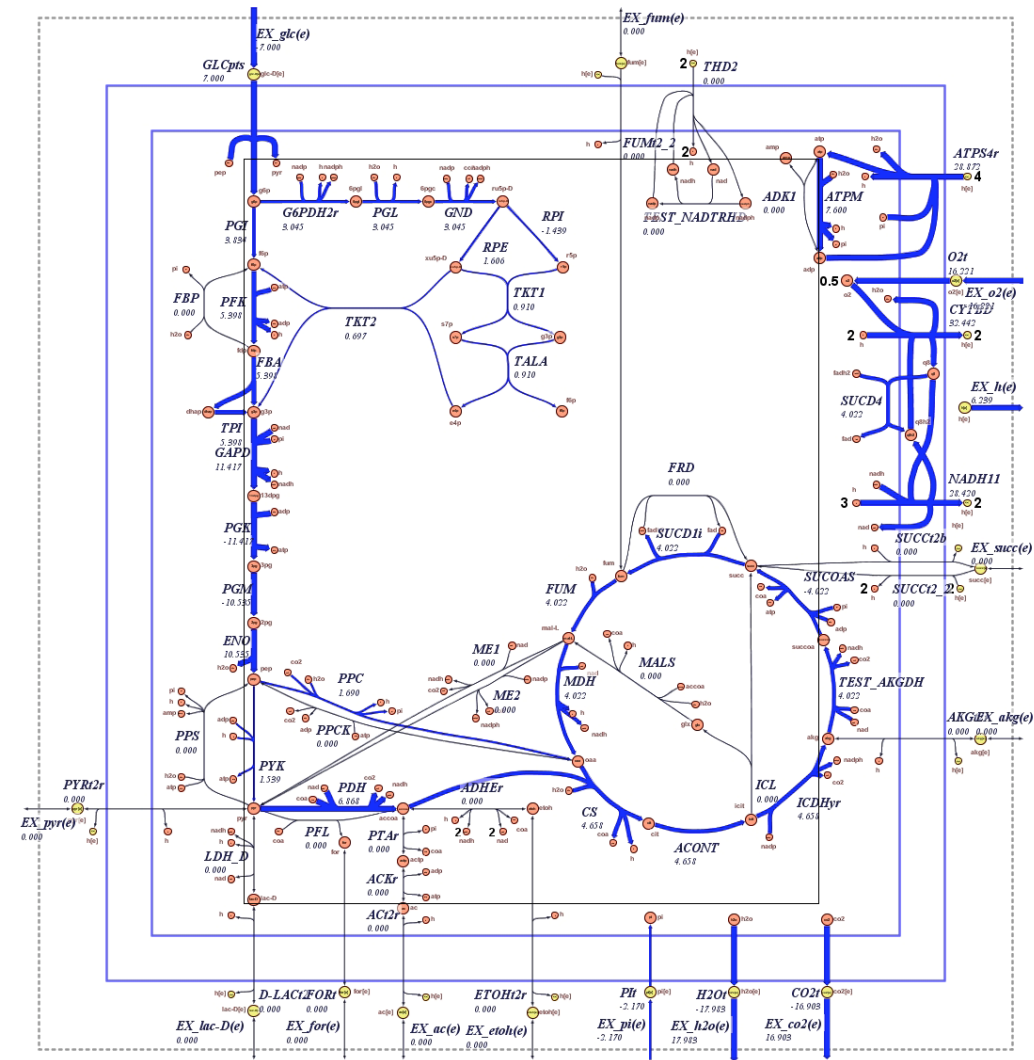




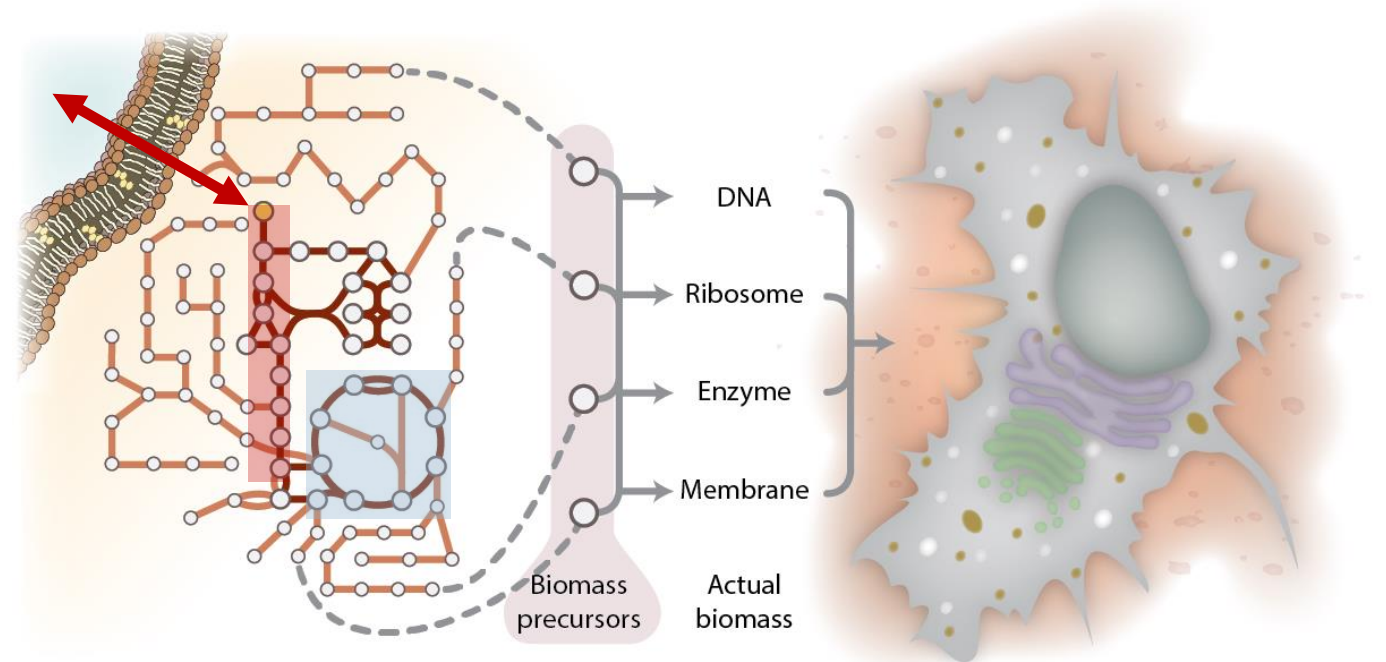
Goals:

High cell concentration and viability
High specific productivity
High/maintain product quality

A METABOLIC MAP RESOURCE THAT ALLOWS YOU TO SIMULATE



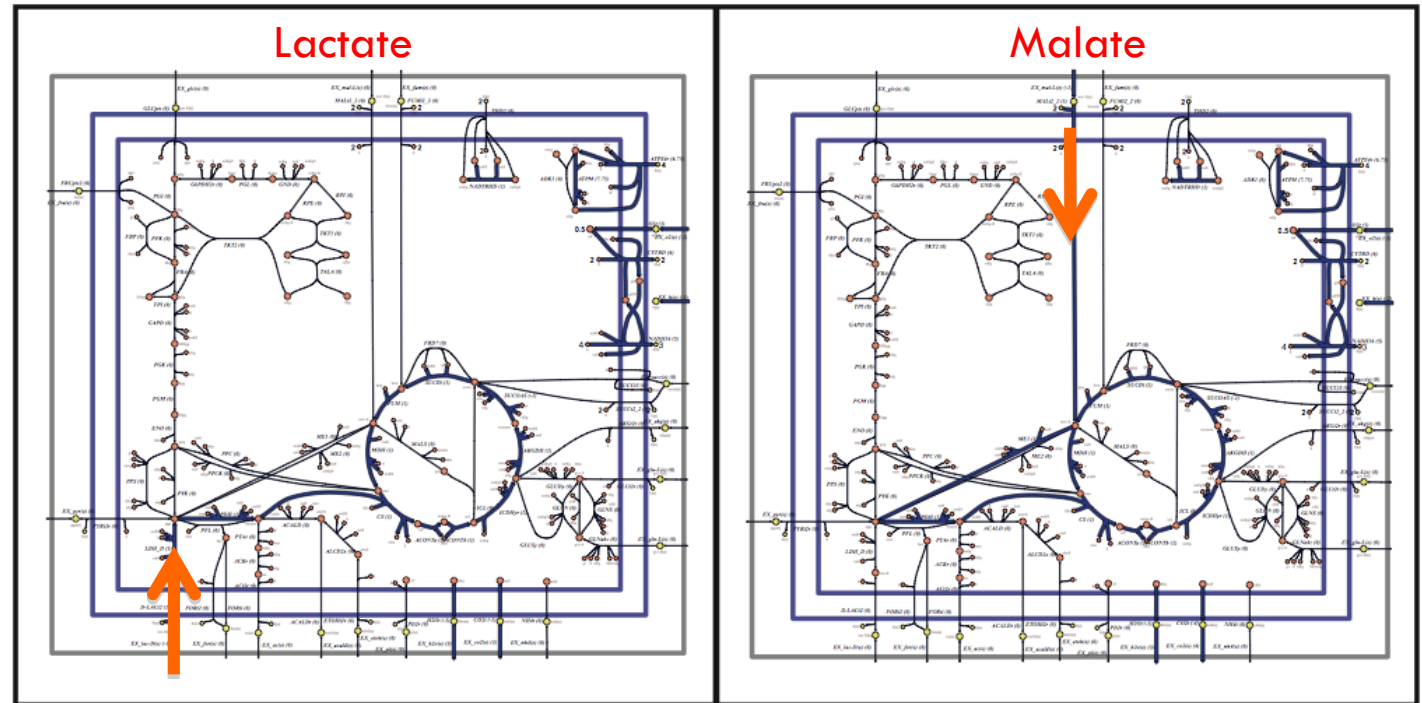
Exo-metabolomics
(spent media analysis)
- Maximum metabolic fluxes in cells

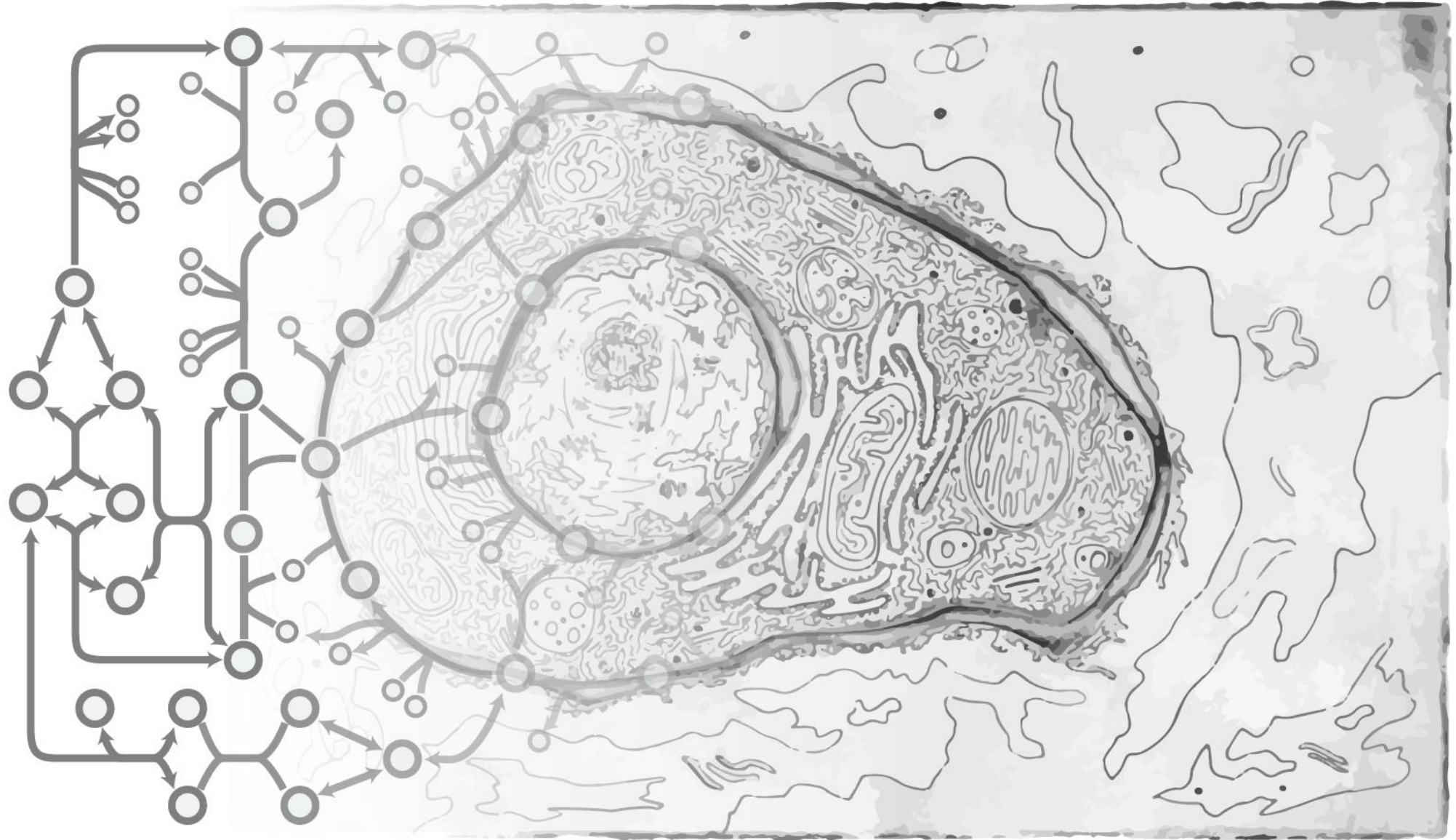


Transcriptomics, Proteomics, etc.
- Expression level of pathways

Endo-metabolomics
- Metabolite changes in cell,
may not relate to flux
- May impact regulation

SIMULATING METABOLIC FLUXES UNDER DIFFERENT GROWTH CONDITIONS





WHERE CAN YOU FIND THE CHO METABOLIC MODEL?

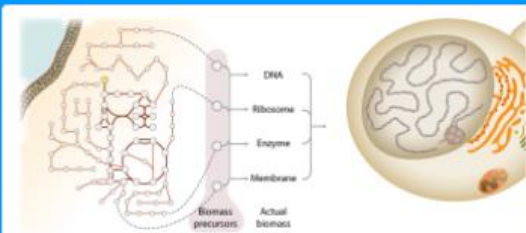
[HTTP://BIGG.UCSD.EDU](http://bigg.ucsd.edu) – A RESOURCE FOR SEARCHING THE CHO METABOLIC PATHWAYS

BiGG Models

Search the database by model, reaction, metabolite, or gene ?

Search

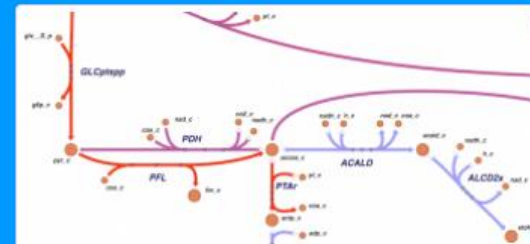
Latest update Version 1.5: Introducing Recon3D



View Models



View Metabolites



View Reactions



General Info ▾ Genomes ▾ Resources ▾ Partners

Genomic Data Resources

This web page contains links to relevant publications and public data sets. Links to next generation sequencing raw data in the Sequence Read Archive (SRA) are provided when available. Journal access may be limited to subscribed users.

[Click here](#) to access the original CH and CHO genome data files that are hosted in our own *Cricetulus griseus* File Archive.

Genome

Description	Title	Reference
CHO cell genome-scale metabolism model with 1,766 genes, 6,663 reactions, and integrated -omics data findings	A Consensus Genome-scale Reconstruction of Chinese Hamster Ovary Cell Metabolism	Hefzi H et al. <i>Cell Systems</i> (2016) 3, 434-443. Zipped CHO, CHO-K1, CHO-S, and CHO-DG44 cell line-specific models [.ZIP file]
A chromosome sorting approach was used to facilitate genome assembly from short-read sequences of the Chinese hamster genome	Chinese hamster genome sequenced from sorted chromosomes	Brinkrolf K et al. <i>Nature Biotechnol</i> (2013) 31, 694-695
Draft genomic sequence of the Chinese hamster and resequencing of CHO-K1, DG44, and CHO-K cell lines	Genomic landscapes of Chinese hamster ovary cell lines as revealed by the <i>Cricetulus griseus</i> draft genome	Lewis N et al. <i>Nature Biotechnol</i> (2013) 31, 759-765
Generation of a physical chromosome map of the CHO-DG44 cell line.	Construction of BAC-based physical map and analysis of chromosome rearrangement in Chinese hamster ovary cell lines.	Cao Y et al. <i>Biotechnol Bioeng</i> (2012) 109, 1357-1367 CHO BAC library sequences
Draft genomic sequence and analysis of the	The genomic sequence of the Chinese	Xu X et al. <i>Nature Biotechnol</i> (2011) 29, 735-

Draft genomic sequence and analysis of the The genomic sequence of the Chinese Xu X et al. *Nature Biotechnol* (2011) 29, 735–



ELSEVIER

Contents lists available at ScienceDirect

Metabolic Engineering

journal homepage: www.elsevier.com/locate/meteng



Application of a curated genome-scale metabolic model of CHO DG44 to an industrial fed-batch process



Cyrielle Calmels^{a,b}, Andréa McCann^{a,c}, Laetitia Malphettes^a, Mikael Rørdam Andersen^{b,*}

^a Department of Upstream Process Sciences, UCB Pharma, Chemin du Foriest 1, 1420 Braine-l'Alleud, Belgium

^b Department of Biotechnology and Biomedicine, Technical University of Denmark, Søltofts Plads 223, 2800 Kgs. Lyngby, Denmark

^c Mass Spectrometry Laboratory, University of Liège, Allée du six aout, Liège, Belgium



Cell Systems

Volume 4, Issue 5, 24 May 2017, Pages 530–542.e6



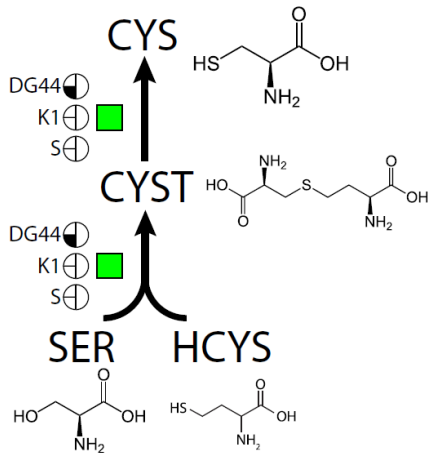
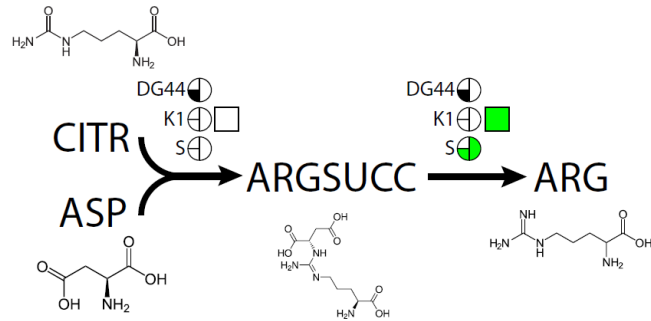
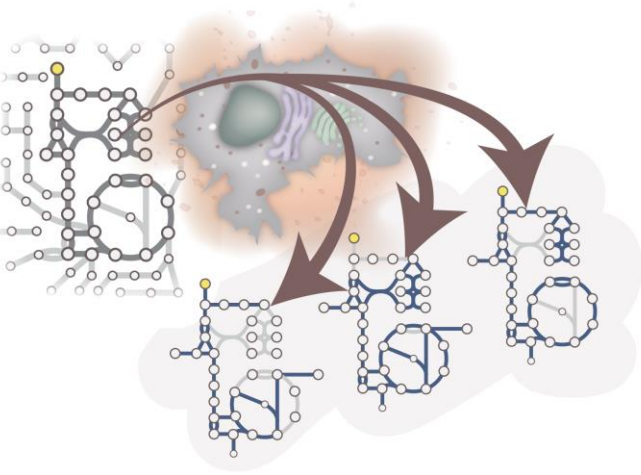
Article

Mammalian Systems Biotechnology Reveals Global Cellular Adaptations in a Recombinant CHO Cell Line

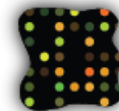
Faraaz Noor Khan Yusufi^{1,6}, Meiyappan Lakshmanan^{1,6}, Ying Swan Ho^{1,6}, Bernard Liat Wen Loo¹, Pramila Ariyaratne², Yuansheng Yang¹, Say Kong Ng¹, Tessa Rui Min Tan¹, Hock Chuan Yeo^{1,3}, Hsueh Lee Lim¹, Sze Wai Ng¹, Ai Ping Hiu¹, Chung Ping Chow¹, Corrine Wan¹, Shuwen Chen¹, Gavin Teo¹, Gao Song², Ju Xin Chin¹ ... Dong-Yup Lee^{1,3,7}  

CELL LINE SPECIFIC MODELS

Cell line specific models



Expression data

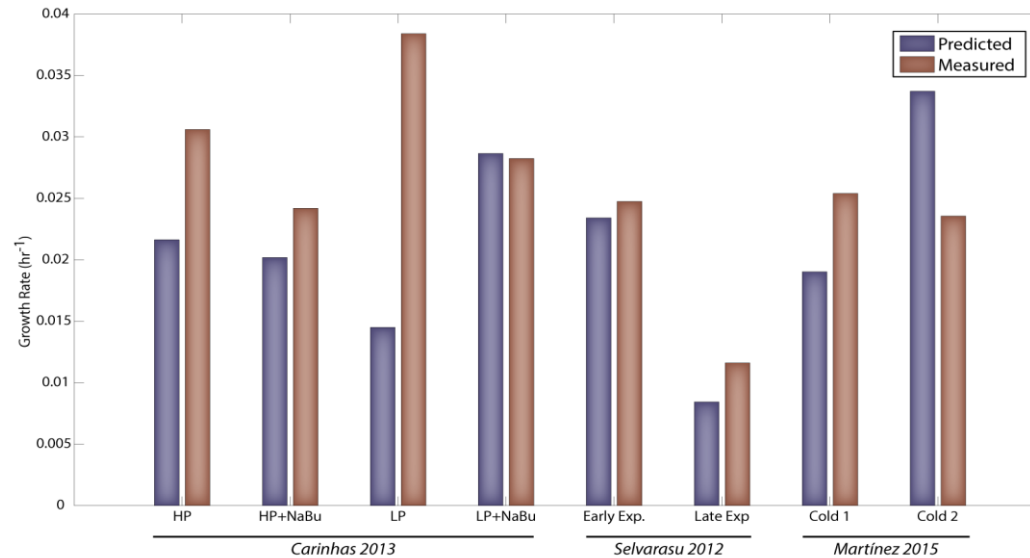


i. Identify reactions with expressed genes

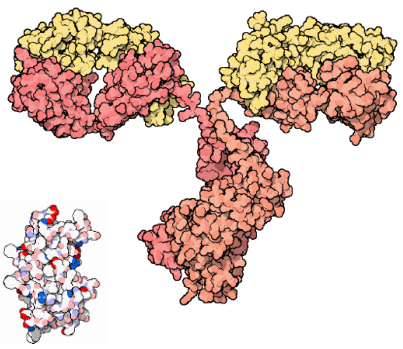
Phenotype data



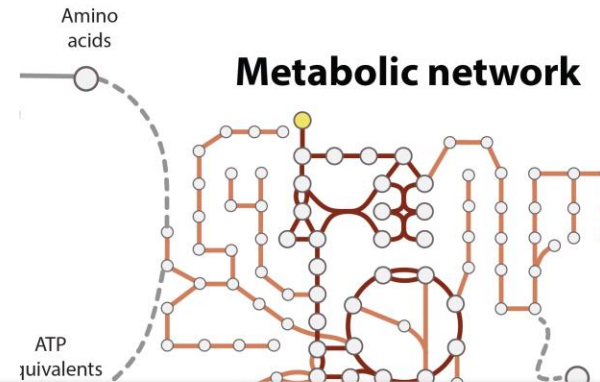
ii. Minimize addition of reactions with low expression and compute consistency score



Opdam, et al., Cell Systems, 2017
 Richelle, et al. bioRxiv, 2018
 Richelle, et al. PLoS Comp Bio, 2019
 Joshi, bioRxiv, 2019
 Laurent, et al. Nat Protocols, 2019
 Hefzi, et al., Cell Systems, 2016



AN EXTENDED FRAMEWORK FOR MODELING PROTEIN SECRETION



Jahir Gutierrez



Amir Feizi



Cold
Spring
Harbor
Laboratory

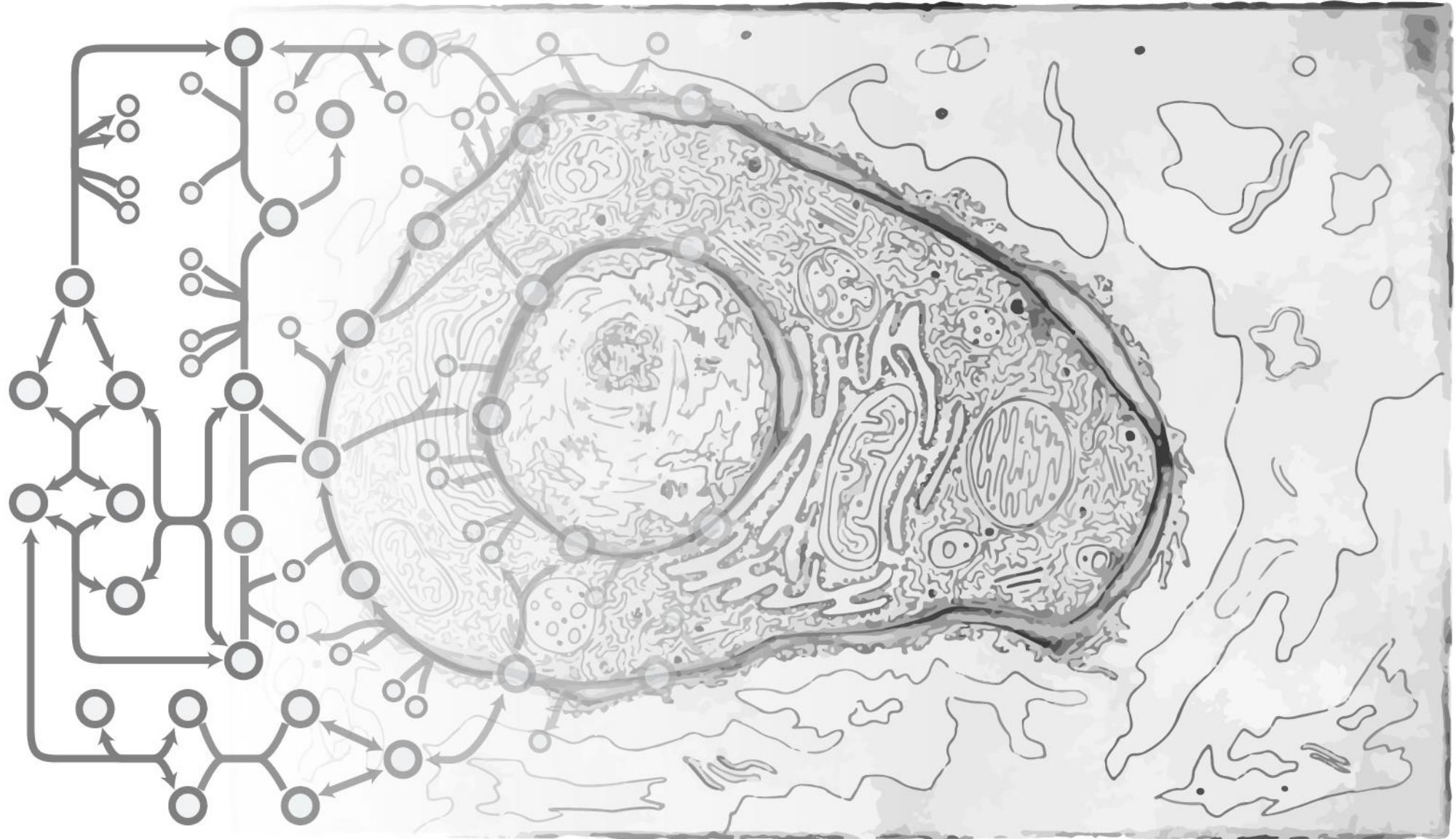
bioRxiv

Genome-scale reconstructions of the mammalian secretory pathway predict metabolic costs and limitations of protein secretion

Jahir M. Gutierrez, Amir Feizi, Shangzhong Li, Thomas B. Kallehauge, Hooman Hefzi, Lise M. Grav, Daniel Ley, Deniz Baycin Hizal, Michael J. Betenbaugh, Bjorn Voldborg, Helene Faustrup Kildegaard, Gyun Min Lee, Bernhard O. Palsson, Jens Nielsen, Nathan E. Lewis

261 machinery proteins

Product-specific needs: Nutrients and machinery



HOW DO I USED A CHO METABOLIC MODEL?

openCOBRA

https://opencobra.github.io

AppsVirtual EMSSave to Mendeley

openCOBRA

COntstraint-Based Reconstruction and Analysis

openCOBRA

Open-source, community-developed code base for COntstraint-Based Reconstruction and Analysis.

RUNNING SIMULATIONS:


COBRA TOOLBOX (MATLAB) Packages

COBRAPY (PYTHON)

ALSO:


SYBIL (R)

OPTFLUX (JAVA)




The COBRA Toolbox

The COntstraint-Based Reconstruction and Analysis (COBRA) Toolbox written in MATLAB.



COBRAPY

COBRAPY is a package for constraint-based modeling of biological networks written in Python.



COBRA.jl

High-level, high-performance, constraint-based reconstruction and analysis in Julia.

GETTING
STARTED

What is flux balance analysis?

Jeffrey D Orth, Ines Thiele & Bernhard Ø Palsson

Flux balance analysis is a mathematical approach for analyzing the flow of metabolites through a metabolic network. This primer covers the theoretical basis of the approach, several practical examples and a software toolbox for performing the calculations.

nature
protocols

PROTOCOL UPDATE

<https://doi.org/10.1038/s41596-018-0098-2>

Creation and analysis of biochemical constraint-based models using the COBRA Toolbox v.3.0

Laurent Heirendt^{1,24}, Sylvain Arreckx^{1,24}, Thomas Pfau², Sebastián N. Mendoza^{3,4}, Anne Richelle⁵, Almut Heinken¹, Hulda S. Haraldsdóttir¹, Jacek Wachowiak¹, Sarah M. Keating⁶, Vanja Vlasov¹, Stefania Magnúsdóttir¹, Chiam Yu Ng⁷, German Preciat¹, Alise Žagare¹, Siu H. J. Chan⁷, Maike K. Aurich¹, Catherine M. Clancy¹, Jennifer Modamio¹, John T. Sauls⁸, Alberto Noronha¹, Aarash Bordbar⁹, Benjamin Cousins¹⁰, Diana C. El Assal¹, Luis V. Valcarcel¹¹, Iñigo Apaolaza¹¹, Susan Ghaderi¹, Masoud Ahookhosh¹, Marouen Ben Guebila¹, Andrejs Kostromins¹², Nicolas Sompairac¹³, Hoai M. Le¹, Ding Ma¹⁴, Yuekai Sun¹⁵, Lin Wang⁷, James T. Yurkovich¹⁶, Miguel A. P. Oliveira¹, Phan T. Vuong¹, Lemmer P. El Assal¹, Inna Kuperstein¹³, Andrei Zinovyev¹³, H. Scott Hinton¹⁷, William A. Bryant¹⁸, Francisco J. Aragón Artacho¹⁹, Francisco J. Planes¹¹, Egils Stalidzans¹², Alejandro Maass^{3,4}, Santosh Vempala¹⁰, Michael Hucka²⁰, Michael A. Saunders¹⁴, Costas D. Maranas⁷, Nathan E. Lewis^{5,21}, Thomas Sauter², Bernhard Ø. Palsson^{16,22}, Ines Thiele¹ and Ronan M. T. Fleming^{1,23*}

Received: 3 October 2016 | Revised: 7 June 2017 | Accepted: 12 July 2017
DOI: 10.1002/bit.26384

REVIEW

WILEY BIOTECHNOLOGY
BIOENGINEERING

Quantitative intracellular flux modeling and applications in biotherapeutic development and production using CHO cell cultures

Zhuangrong Huang¹ | Dong-Yup Lee^{2,3} | Seongkyu Yoon¹



The COBRA Toolbox

[Search docs](#)

[Home](#)

Installation

Functions

Tutorials

Analysis

Base

Data integration

Design

Reconstruction

Visualization

How to contribute

How to cite

Support

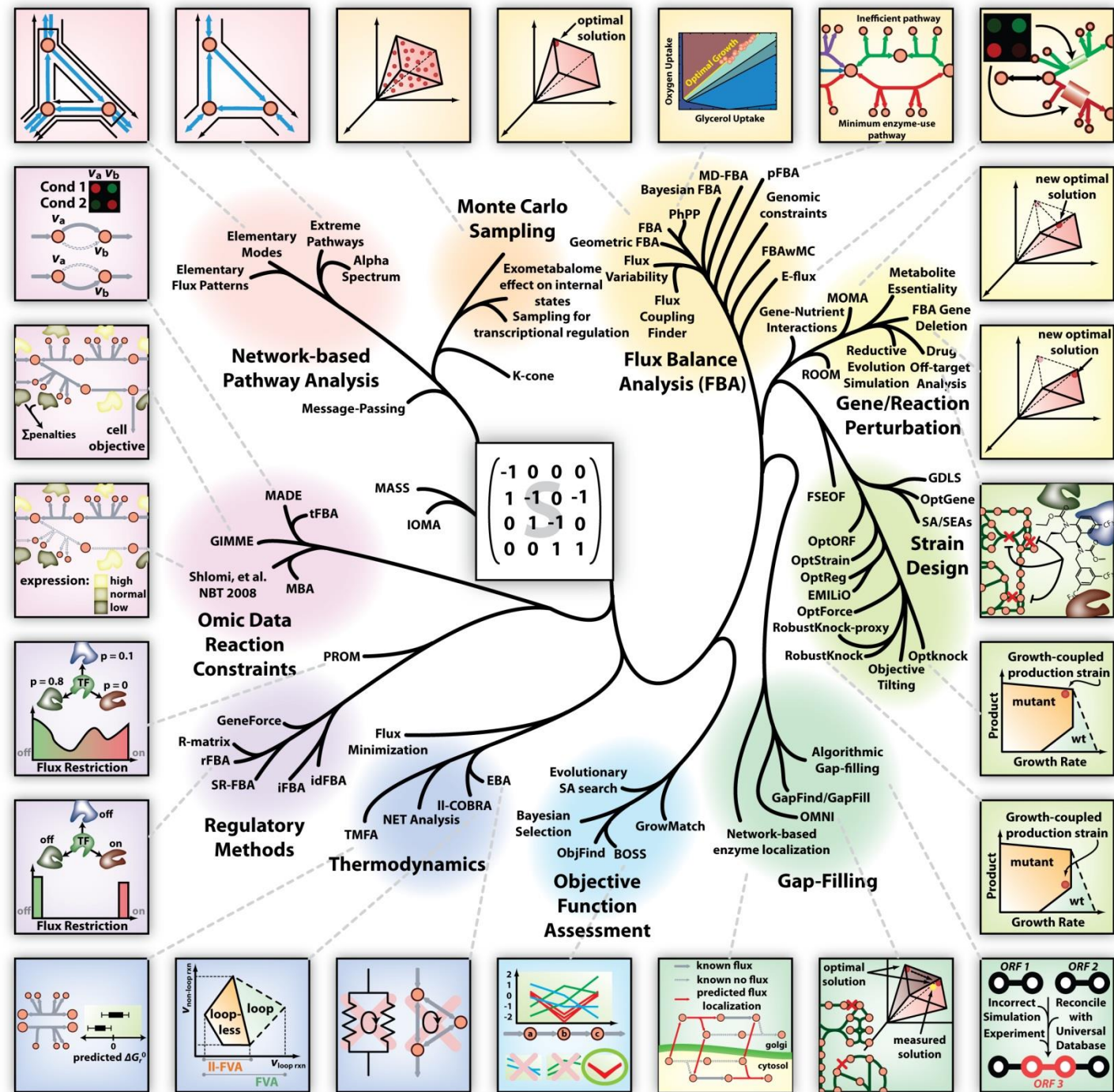
[Docs](#) » [Tutorials](#)

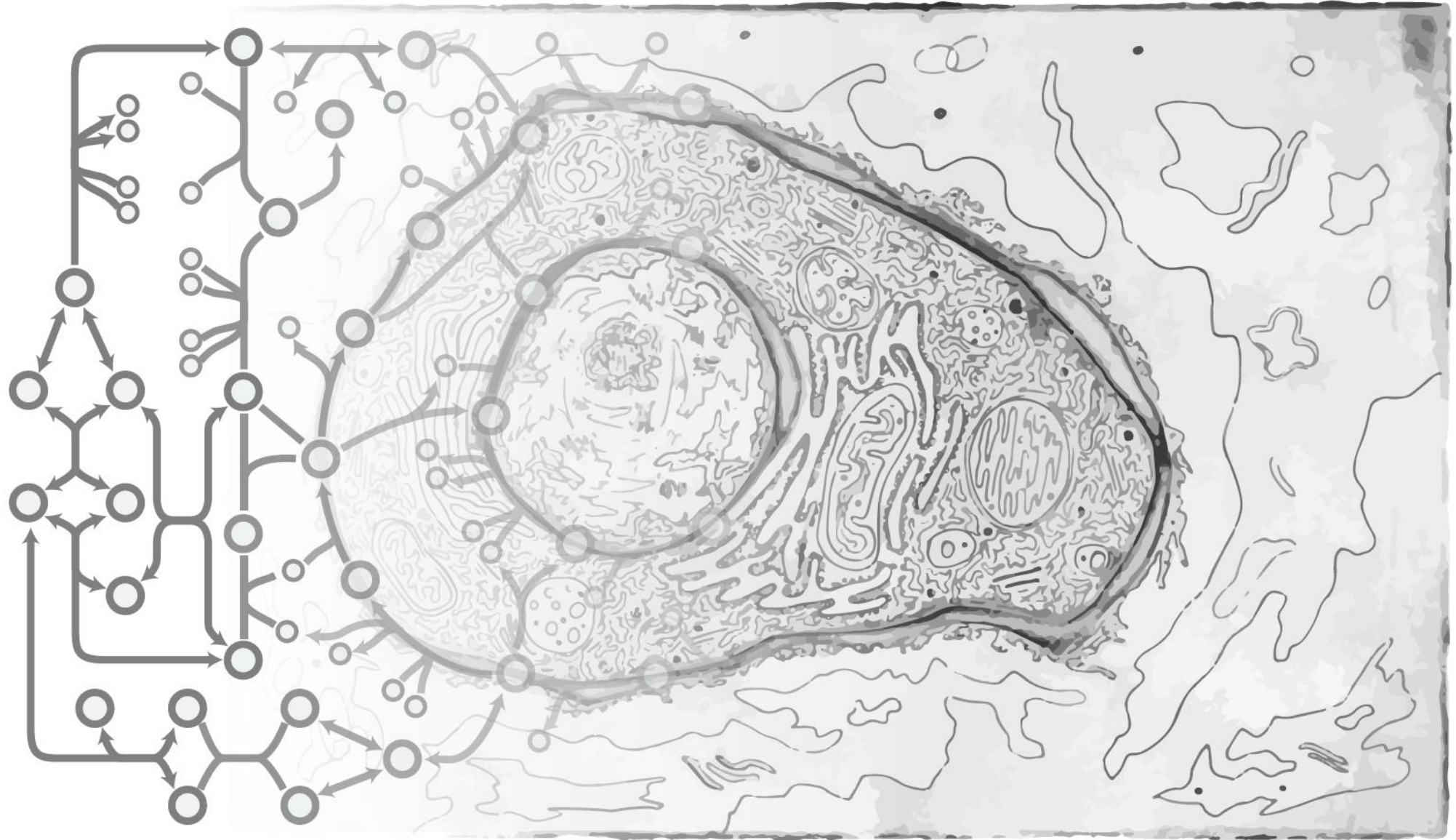
Tutorials

Analysis

- A step-by-step guide to parsimonious enzyme usage Flux Balance Analysis - pFBA
- Analyze Steady-State Community COBRA Models
- Atomically resolve a metabolic reconstruction
- Browse Networks in the Matlab Command Window Using surfNet
- Computation and analysis of microbe-microbe metabolic interactions
- Computation and analysis of rescued lethal gene deletions in a host-microbe model
- Creation and simulation of personalized microbiota models through metagenomic data integration
- Determining MinSpan vectors of COBRA model
- Flux Balance Analysis (FBA)
- Flux Variability analysis (FVA)
- genetic Minimal Cut Sets - gMCS
- Proton shuttle testing with sparse flux balance analysis
- Reaction essentiality across multiple models
- Relaxed Flux Balance Analysis: Recon 3
- Relaxed Flux Balance Analysis: Toy model
- Simulation of growth of human gut microbes on different diets
- Sparse Flux Balance Analysis
- Sparse flux balance analysis test for a minimal stoichiometrically balanced cycle involving ATP

The toolbox of constraint-based methods for computational modeling goes far beyond FBA... now >200 methods





HOW DO I VISUALIZE SIMULATIONS OR
DATA ON A METABOLIC MODEL?

[GitHub](#) [Docs](#) [What's new?](#)



ESCHER

Build, share, and embed visualizations of metabolic pathways

Filter by organism

All ▾

Map

Glycolysis TCA PPP (RECON1) ▾

Model (Optional)

RECON1 ▾

Tool

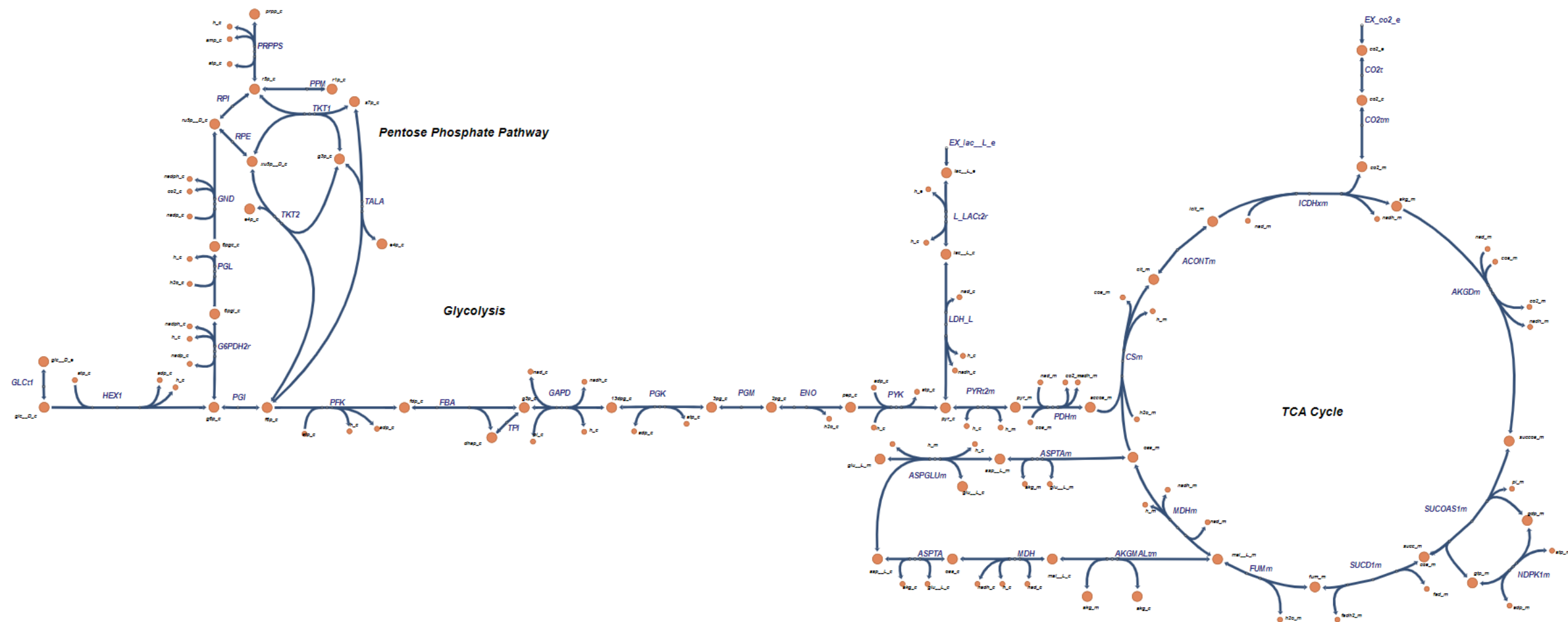
Builder ▾

Options

- ☐ Scroll to zoom (instead of scroll to pan)
- ☐ Never ask before reloading

Load map

Map ▾ Model ▾ Data ▾ Edit ▾ View ▾ ?



Button bar

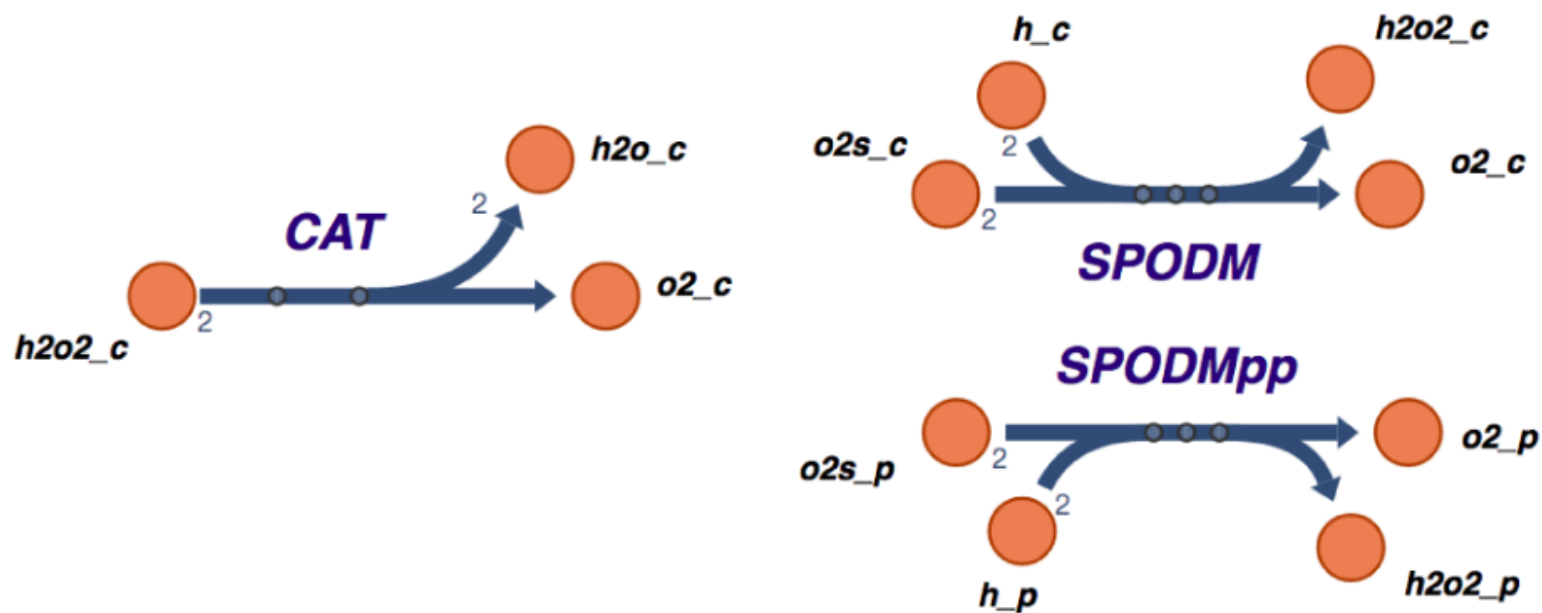
Menus

Map ▾ Model ▾ Data ▾ Edit ▾ View ▾

?

Documentation

Oxidative Stress



— Jump to map —

Jump to map menu

The metabolic map illustrates the TCA cycle and its associated pathways in *E. coli*. The cycle is represented by a central loop of nodes (circles) connected by arrows. The nodes are labeled with gene names (e.g., *ENO*, *GLCptspp*, *PTAr*, *ACKr*, *ACALD*, *ACT2rpp*, *ACTex*, *ETOHtrpp*, *ETOHtex*) and their corresponding protein names (e.g., *ENO 37.9*, *GLCptspp 20.0*, *PTAr 15.9*, *ACKr -15.9*, *ACALD -15.9*, *ACT2rpp -16.2*, *ACTex -16.2*, *ETOHtrpp -15.9*, *ETOHtex -15.9*). The map also shows the entry of pyruvate into the cycle via the *PYR* gene and the exit of the cycle via the *GLCptspp* gene. The map is color-coded: red for the main TCA cycle, purple for the *GLCptspp* pathway, and blue for the *ETOH* pathway. The map is titled "Metabolic map of the TCA cycle and related pathways in *E. coli*".

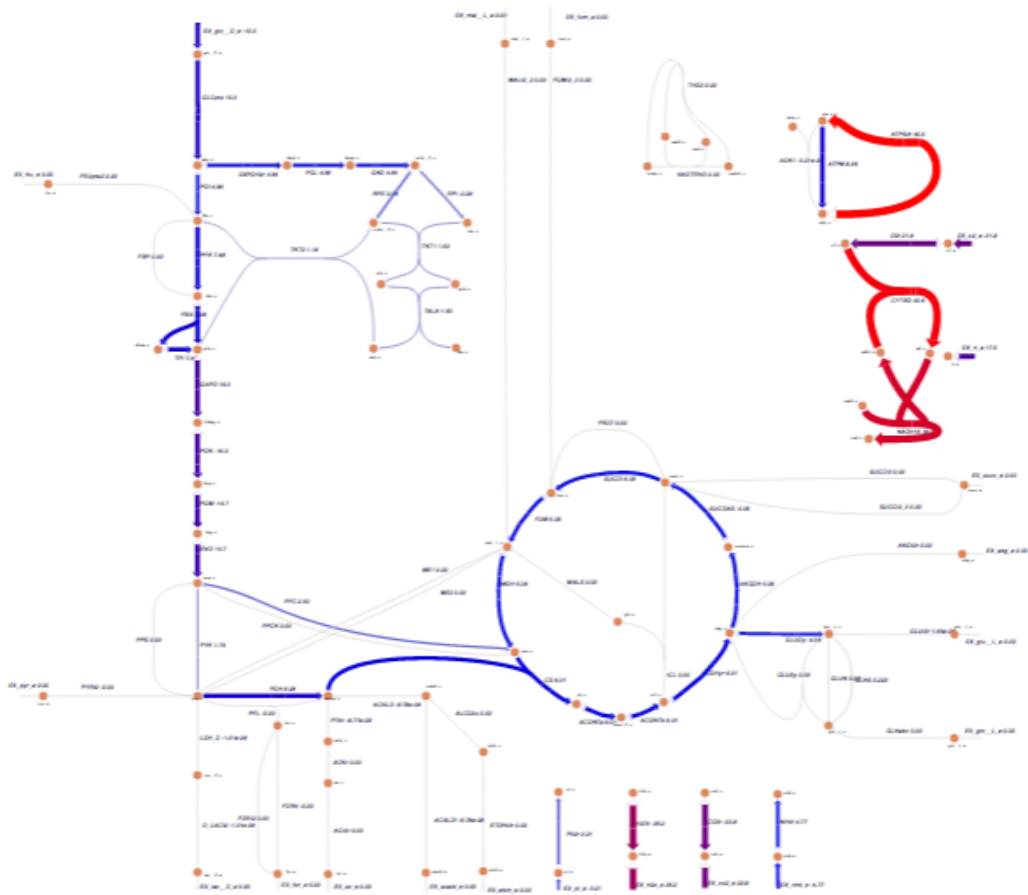
Settings
✓ Hide secondary metabolites
✓ Absolute value for reactions

- ✓ Hide secondary metabolites
- ✓ Absolute value for reactions

ESCHER AND IPYTHON

```
In [10]: b = escher.Builder(map_name='e_coli_core.Core metabolism',
                             reaction_data=solution.x_dict,
                             # color and size according to the absolute value
                             reaction_styles=['color', 'size', 'abs', 'text'],
                             # change the default colors
                             reaction_scale=[{'type': 'min', 'color': '#cccccc', 'size': 4},
                                              {'type': 'mean', 'color': '#0000dd', 'size': 20},
                                              {'type': 'max', 'color': '#ff0000', 'size': 40}],
                             # only show the primary metabolites
                             hide_secondary_metabolites=True)
b.display_in_notebook()
```

Out[10]:



SUMMARY

Metabolic modeling elucidates the inner workings of metabolism in a cell

Genome-scale metabolic models exist for the Chinese hamster and others

- <http://CHOgenome.org/>
- <http://bigg.ucsd.edu/>
- <https://www.metanetx.org/>

Models can be trimmed to focus on cell lines (or even subsets of pathways)

Simulation tools exist in several languages: Matlab, Python, R, Java, Julia, etc.

Visualization tools can be deployed using web-based tools or others

- <http://escher.github.io/>

These slides can be downloaded at <http://lewislab.ucsd.edu>